

Cardiac allograft vasculopathy: a review

Danny Ramzy, MD;* Vivek Rao, MD, PhD;* Julie Brahm, BSc;* Santiago Miriuka, MD;†
Diego Delgado, MD;† Heather J. Ross, MD, MSc†

Cardiac allograft vasculopathy (CAV) is a major factor limiting long-term survival after cardiac transplantation. CAV is an accelerated form of coronary artery disease (CAD) that is characterized by concentric fibrous intimal hyperplasia along the length of coronary vessels. Both immunologic and non-immunologic risk factors contribute to the development of CAV by causing endothelial dysfunction and injury eventually leading to progressive intimal thickening. The diagnosis of CAV remains a challenge as angiography, the standard method for detecting focal plaques, lacks sensitivity in detecting CAV, and intravascular ultrasonography, a more sensitive method, lacks the ability to evaluate the entire coronary tree. The disease is difficult to treat and results in significant morbidity and mortality. Since treatment of CAV is limited and usually involves repeat transplantation, prevention or mitigation of immunologic and nonimmunologic risk factors is critically important. CAV prevention may involve therapy that provides protection against endothelial injury implemented just before transplantation, during storage and transplantation as well as after transplantation. This review addresses the frequency of occurrence, pathophysiology, diagnosis and treatment of CAV, highlighting areas of active research.

La vasculopathie de l'allogreffe cardiaque (VAC) est un facteur important qui limite la survie à long terme après une transplantation cardiaque. La VAC est une forme accélérée de coronaropathie caractérisée par une hyperplasie fibreuse concentrique de la tunique interne des vaisseaux coronariens. Des facteurs de risque immunologiques et non immunologiques contribuent à l'apparition de la VAC en causant une dysfonction de l'endothélium et une lésion qui entraîne éventuellement l'épaississement progressif de la tunique interne. La VAC est toujours difficile à diagnostiquer, car l'angiographie, méthode normalisée de détection de la plaque focale, n'est pas assez sensible pour détecter la VAC; l'échographie intravasculaire, plus sensible, ne permet pas d'évaluer l'arbre coronarien au complet. La maladie est difficile à traiter et entraîne un taux important de morbidité et de mortalité. Comme le traitement de la VAC est limité et entraîne habituellement une nouvelle transplantation, il est crucial de prévenir ou d'atténuer les facteurs de risque immunologiques et non immunologiques. La prévention de la VAC peut faire appel à une thérapie qui protège contre les lésions endothéliales et est mise en œuvre immédiatement avant la transplantation, pendant l'entreposage et la transplantation, ainsi qu'après l'intervention. Cette critique porte sur la fréquence de l'occurrence, la pathophysiologie, le diagnostic et le traitement de la VAC et met en évidence les domaines où des recherches actives sont en cours.

Heart transplantation is the accepted therapy for patients with refractory end-stage heart disease. Although this procedure can extend and improve quality of life, it is not a cure. The median survival after heart transplantation remains 9.3 years, 11.8 years for patients surviving the first year after transplantation.¹ Cardiac allograft vasculopathy (CAV), an accelerated form of coronary artery dis-

ease (CAD), is the leading cause of death between 1 and 3 years after transplantation according to the Registry of the International Society for Heart and Lung Transplantation.¹ After year 3, CAV accounts for 17% of deaths. Angiographic studies indicate that CAV occurs in 42% of all heart transplant patients 3 years after transplantation.¹ Intravascular ultrasonography, a more sensitive technique, de-

tects CAV in 75% of patients at 3 years. Allograft vasculopathy is a phenomenon not limited to cardiac transplantation. A similar process also limits long-term graft survival in other solid organ transplants.

CAV affects arteries, arterioles, capillaries and occasionally veins, with sparing of all recipient vessels.²⁻⁴ The predominant feature of CAV is a diffuse, progressive thickening of the ar-

*From the Heart Transplant Program, Toronto General Hospital, University Health Network, and the Divisions of *Cardiac Surgery and †Cardiology, University of Toronto, Toronto, Ont.*

Accepted for publication Apr. 5, 2005.

Correspondence to: Dr. Vivek Rao, Surgical Director, Cardiac Transplant and Mechanical Circulatory Support, 4N-464, Toronto General Hospital, 200 Elizabeth St., Toronto ON M5G 2C4; fax 416 340-3337; vivek.rao@uhn.on.ca

terial intima that develops in both the epicardial and intramyocardial arteries of the transplanted heart. The process is a concentric fibrous intimal hyperplasia that appears along the entire length of the affected arteries. Included in this form of arteriosclerosis are features of both atherosclerosis and arteritis. The atherosclerotic changes range from a diffuse incorporation of lipids to the development of classic focal plaques later in the disease process. With arteritis, there is thickening of the vessel due to infiltration by mononuclear inflammatory cells responding to alloimmune or infectious stimuli. Rarely, this arteritis may progress to destroy the internal elastic lamina and involve the media.⁵⁻⁸

Differences between cardiac allograft vasculopathy and coronary artery disease

The pathological features of CAV differ significantly from those of CAD (Table 1). CAD is usually a focal, eccentric proliferation of the intima of proximal coronary vessels. There is usually sparing of the intramyocardial vessels. Fatty streaks are seen initially. Of importance in CAD is the deposition of calcium and disruption of the elastic lamina. Rarely there are signs of inflammation, and veins are never involved.

CAV is typically characterized as a diffuse concentric proliferation of the intima. Intramyocardial vessels are usually involved, and the process can even involve the coronary veins. The initial lesions seen are smooth-muscle proliferation of the intima. There is

rarely any calcium deposition,⁵ the internal elastic lamina is intact and inflammation is usually present.

Pathological features

CAV is mainly a disease of the intima. Changes in the intima can be seen as early as 6 months after transplantation. The lesion at this time is a mild intimal thickening. Mild fibrosis and increases in extracellular matrix proteins may be present. Early after transplantation, intimal thickening is limited to the proximal arteries.⁹ These lesions are characterized by hyperplastic fibrous thickening. Lesions then progress to a fibrofatty atheromatous plaque. Ultimately, the coronary vasculature progresses to a diffuse fibrous thickening of the intima, which can have superimposing atheromatous plaques.

The internal elastic lamina is almost always intact except for breaks that may be seen in more advanced stages of CAV. The media can be unaffected or almost completely replaced by fibrous tissue. As the intimal disease progresses in severity so does fibrosis of the media. The only vessels relatively unaffected are those with little or no muscular layer.

Pathophysiologic characteristics

The pathophysiologic features of CAV, although not completely understood, likely involve components of both immune-mediated and non-immune-mediated endothelial damage, and passenger “native vessel”

atherosclerosis.¹⁰ There is substantial evidence that immunologic factors, including histocompatibility mismatch, acute rejection episodes and chronic inflammation, play a major role in CAV development. Nonimmunologic factors include cause of donor brain death, cytomegalovirus (CMV) infection, age, sex, obesity, dyslipidemia, hyperhomocysteinemia (HHcy), diabetes mellitus, hypertension, smoking and ischemia-reperfusion injury.¹¹ In general, hyperlipidemia and insulin resistance are the most significant nonimmunologic factors, occurring in 50%–80% of the heart transplant population.¹²

The endothelial damage involved in CAV can be categorized into either denuding or nondenuding injury. In nondenuding injury a rapid replacement of injured endothelial cells leads to endothelial dysfunction.⁸ Both immune-related and nonimmune-related factors contribute to nondenuding injury. In contrast, denuding injury is caused by ischemia-reperfusion injury during transplantation or during episodes of acute cellular rejection. This results in the loss of large stretches of endothelium along the vessel, which causes significant endothelial dysfunction.¹³ According to one hypothesis, it is the immune component or alloantigen-dependent mechanism of injury that acts principally to intensify initial nonimmune damage to the endothelial cells.¹⁴ Denuding injury allows for blood components and circulating cytokines to have direct contact with the subintimal layers. This can lead to significant proliferation of smooth-muscle cells. Therefore, CAV can be initiated or exacerbated by several processes that can lead to denuding or nondenuding injury. These include ischemia-reperfusion injury, immune activation, viral infection and injury from immunosuppressive drugs.

Hyperlipidemia is commonly seen in cardiac transplant patients for several reasons. Many of these patients are hyperlipidemic before transplantation. In addition, the immunosup-

Table 1
Comparison between cardiac allograft vasculopathy and coronary artery disease

Characteristic	Cardiac allograft vasculopathy	Coronary artery disease
Vessel involvement	All vessel types within the allograft Mostly intramyocardial vessels	Proximal coronary vessels
Plaque pattern	Diffuse and concentric	Focal and eccentric
Inflammation	Yes	Rarely
Internal elastic lamina	Intact	Disrupted
Calcium deposition	No	Yes

pressive therapy given to patients, especially calcineurin inhibitors, may result in or exacerbate pre-existing dyslipidemia. Hypercholesterolemia, in a rabbit heterotopic cardiac transplant model, has been shown to be associated with CAV^{15,16} and transplanted coronary arteries were more affected by hypercholesterolemia than native coronary arteries. Hypercholesterolemia promotes fibrofatty proliferative changes to the intimal hyperplasia seen in most patients with CAV.¹⁵

In solid-organ transplant recipients, HHcy is extremely common and occurs early with a rate as high as 80%–90%.^{17–28} HHcy can damage cells by several mechanisms, but primarily by affecting the endothelium.^{29–31} HHcy results in reduced endothelial nitric oxide production,^{32,33} impaired arterial response to vasodilators and increased expression of procoagulant factors.^{22,29–31} The neutrophil–endothelium interaction is promoted in the setting of HHcy, allowing for the presence of more neutrophils in the intima. All of these alterations in the endothelial wall are caused by alterations in the redox state induced by high homocysteine levels.^{30–32} Several investigators have demonstrated that HHcy is associated with the development of CAV.^{34,35}

Hypertension, smoking, diabetes mellitus and other risk factors for atherosclerosis are associated with CAV. Hypertension in transplant patients can be present preoperatively or postoperatively secondary to immunosuppressive medication, such as cyclosporine. Hypertension causes endothelial injury by promoting the formation of intimal hyperplasia, which eventually gives rise to atherosclerotic lesions.

Although the relative importance of the direct versus the indirect pathway of alloreactivity is still debated, one theory is that direct activation of recipient CD4+ T cells by donor allograft/nonsel self major histocompatibility complex (MHC) class II molecules initiates graft rejection. CD8+ T cells can become activated by previously activated CD4+ T cells

through the CD40L pathway and by nonself MHC class I molecules.

The activation of CD4+ and CD8+ T cells leads to further synthesis of cytokines, which perpetuate the ongoing cascade of events that lead to CAV. The most important cytokines in allograft rejection are interleukin-2 (IL-2), interferon-gamma (IFN- γ) and tumour necrosis factor-alpha (TNF- α). IL-2 induces T-cell proliferation and differentiation, IFN- γ activates macrophages, and TNF- α itself is cytotoxic to the transplanted heart. In addition, TNF- α acts to increase MHC class I expression, while IFN- γ increases the expression of both MHC classes I and II molecules. Overall, these cytokines can lead to chronic graft rejection. IFN- γ and TNF- α also induce the leukocyte vascular cell adhesion molecule-1, which promotes monocyte adhesion and entry through the endothelium, leading to CAV.

Ardehali and associates³⁶ used a murine CAV model with a compromised indirect alloreactivity pathway to show that this did not affect the extent of intimal thickening or lymphocyte and macrophage infiltration after heart transplantation when compared with wild type mice. They proposed 2 potential explanations: (1) an impaired indirect pathway is enough to cause severe CAV and the direct pathway does not play a major role in CAV; or (2) the direct pathway of alloreactivity can fully compensate for the impaired indirect pathway.³⁶ Other studies, such as the one by Game and associates,³⁷ have found a correlation between increased indirectly activated T cells and chronic rejection. As far as the importance of CD4+ versus CD8+ T cells, it appears that the CD4+ allorecognition pathway is required for CAV development, whereas the CD8+ pathway may act to increase the severity of CAV.³⁸ In a study by Szeto and colleagues,³⁹ hearts transplanted into CD8+ T-cell-depleted rats developed CAV, but there was no CAV in the CD4+ T-cell-depleted recipient.

These findings suggest that CAV is dependent on CD4+ indirect allorecognition and not a CD8+ direct pathway. It remains controversial as to which component of the immune response is involved in CAV, but most transplant centres agree that immune activation plays a role.

Acute rejection as a cause or risk factor for CAV has been investigated by several authors.^{40–43} Some groups have reported an association between the severity and frequency of rejection and the severity of CAV; however, others have reported that episodes of acute rejection are not associated with the development of CAV.^{40–44} One proposed mechanism linking acute rejection to CAV is that the inflammatory process and tissue destruction from rejection result in endothelial damage, which initiates the process of CAV or potentiates the CAV already in progress.

Recent research has correlated ischemia–reperfusion injury with CAV. Determinants of ischemia–reperfusion injury are length of ischemic time and methods of allograft storage. Gohra and associates⁴⁵ demonstrated in a rodent model of orthotopic aortic allograft transplantation that ischemia and reperfusion result in endothelial injury, leading to the development of transplant vasculopathy. They also found that endothelial cell loss occurred in both isografts and allografts due to ischemia and reperfusion.⁴⁵ This initial loss of endothelial cells was replaced by 2 weeks; however, transplant vasculopathy developed within 60 days.⁴⁵ Their study indicated that ischemia and reperfusion injury led to the development of transplant vasculopathy since isografts developed vasculopathy, although to a lesser extent than the allografts.

Several changes occur to the endothelium following hypoxia, including loss of the ability to release nitric oxide within minutes after reperfusion.^{45–48} This loss is related to the consumption of nitric oxide by superoxide radicals formed early during reperfusion.⁴⁹ Ex-

perimental evidence suggests that the oxygen free radicals are produced by neutrophils.⁵⁰ In vitro exposure of coronary arteries to oxygen radicals produces endothelial dysfunction.⁵⁰ In addition, ischemia–reperfusion causes the endothelial cells to become activated and express surface adhesion molecules. These molecules promote circulating leukocyte adhesion, which then causes endothelial damage by direct cytotoxicity.^{51–53} These leukocytes become activated and release cytokines, which enhance leukocyte and smooth-muscle cell proliferation and activation. Ischemia also promotes complement activation that causes not only cell lysis, but results in several other changes such as increased vessel permeability, leukocyte chemoattraction and smooth-muscle contraction.^{54,55} Complement I-q can increase platelet procoagulant activity, which can enhance CAV by the formation of thrombus but mostly by causing the release of several vasoactive substances and growth factors such as platelet-derived growth factor-beta, thromboxane A and prostacyclin.⁵⁴ These are few of the mechanisms by which ischemia–reperfusion initiates the process of CAV. In a recent study, myocardial ischemia complicated by fibrosis in the peritransplant period was associated with increased progression of CAV and a poorer long-term outcome.⁵⁶

Several investigators have reported an association between pathogens (*Chlamydia pneumoniae*, CMV, herpes simplex, parvovirus) and CAV. Subramanian and colleagues⁵⁷ have demonstrated that *C. pneumoniae* infection is correlated with the severity of CAV. They concluded that CAV developed in heart transplant recipients who tested positive for immunoglobulin-G against *C. pneumoniae* but not in those who tested positive for *C. pneumoniae* by polymerase chain reaction.⁵⁷ Again, this finding implicates an immunologic mechanism behind the development of CAV, regardless of the inciting stimulus. CMV infection has been as-

sociated with both atherosclerosis and CAV. The Stanford group demonstrated that severe CAV developed in approximately 30% of CMV-infected heart transplant recipients, representing a 3-fold increase compared with uninfected recipients.⁵⁸ CMV has the ability to infect vascular endothelial cells and induce endothelial injury, which can lead to CAV. Weis and colleagues⁵⁹ reported elevated asymmetric dimethylarginine (a nitric oxide synthase inhibitor) impairing vascular homeostasis in CMV-infected patients. These higher levels can lead to endothelial dysfunction and correlate with increased severity of CAV.⁵⁹ CMV and herpes simplex viruses induce the host adaptive immune response, which leads to the release of cytokines, increased expression of adhesion molecules and activation of T-cell responses. Therefore, viral infection may result in CAV by impairing nitric oxide homeostasis, inducing proinflammatory cytokines, and enhancing T-cell-mediated alloreactivity.

The incidence of significant donor CAD remains low, at approximately 2%. Donor CAD can serve as a starting point for CAV and may accelerate the disease process. Donor CAD can be important in the prognosis of the transplant patient in that it can

progress independently of the CAV process. However, Botas and associates⁶⁰ found no significant difference in the rate of intimal thickening between patients with donor hearts having pre-existing coronary artery disease and those without. Thus, the impact of native vessel atherosclerosis on CAV remains controversial.

Finally, cause of donor brain death, more specifically explosive donor brain death (v. gradual brain death), causes an up-regulation of MHC classes I and II antigens, adhesion molecules and cytokine secretion, setting off an accelerated inflammatory response in the heart.^{61–63}

CAV is a complex disease with a multifactorial etiology, and several methods must be adopted to prevent its initiation and progression (Fig. 1).

Diagnosis

Cardiac denervation at the time of heart transplantation usually prevents transplant patients from experiencing angina, which is an important warning sign for heart disease. Only 10%–30% of heart transplant recipients regain any innervation to the heart. Because of this lack of early clinical symptoms, transplant patients with CAV typically present late with silent myocardial infarction, loss of

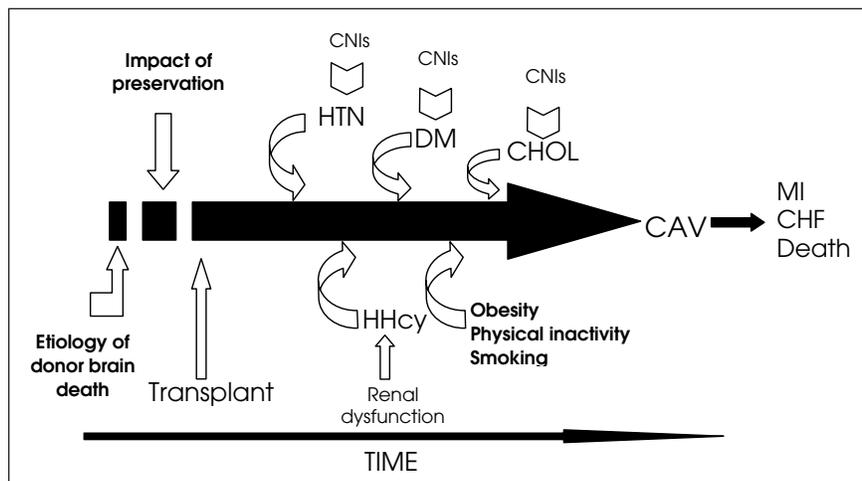


FIG. 1. Pathophysiology of cardiac allograft vasculopathy (CAV). CHF = congestive heart failure, CHOL = cholesterol, CNI = calcineurin inhibitors, DM = diabetes mellitus, HHcy = hyperhomocysteinemia, HTN = hypertension, MI = myocardial infarction.

allograft function or sudden death.⁵

Another difficulty faced by clinicians in diagnosing CAV is coronary remodelling and the diffuse nature of the disease. Angiography measures luminal diameter and compares the narrowing at plaques to normal reference diameters and previous angiograms in order to understand the severity and rate of disease progression. CAV, however, shows no initial decrease in luminal diameter due to vascular remodelling.⁶³ Only when the process is more advanced does the lumen narrow and angiographic detection become possible. Since CAV involves the entire coronary arterial tree, angiography may convey the impression of less-than-actual vessel narrowing at plaque sites. Thus, angiography, although it is a good screening tool for CAD, often underestimates CAV, and in some patients with evenly distributed disease throughout the coronary tree, CAV can be missed altogether.⁶⁴

Despite the poor sensitivity of angiography, it is still widely used as a screening test for vascular disease. Johnson and associates⁹ developed a classification system based on the varying morphologies in CAV to aid in its diagnosis using angiography. Briefly, type A lesions appear as discrete proximal tubular stenoses, type B as diffuse concentric middle or distal stenoses, with type B₁ having an abrupt narrowing and type B₂ having a smooth concentric tapering. Finally a type C angiographic appearance indicates irregular vessels with distal lesions and loss of small branches. Diagnosis of CAV requires type B or C lesions and comparison with previous and recent angiograms to note disease progression.¹⁰

A more sensitive tool is intravascular ultrasonography (IVUS). IVUS is useful for detecting the extent of intimal thickening by imaging vessel wall structure rather than simply luminal diameter. IVUS has an axial resolution of 50–80 µm.⁶³ Unfortunately, it is physically restricted to the larger epicardial arteries, and thus

cannot be used to screen for CAV throughout the entire heart. One year after transplantation, IVUS detects CAV in 50% of patients whereas angiography detects disease in only 10%–20% of patients.^{65,66}

With IVUS, normal coronary intimal thickness ranges between 0.10 and 0.30 mm. Hence, CAV is considered present when intimal thickness exceeds 0.3 mm or when the sum of the intimal and medial thickness exceeds 0.5 mm. At greater than 0.6-mm intimal thickening, patients are 10 times more likely to experience a cardiac event.⁶⁷

Since angiography and IVUS are invasive tests, they pose increased risks for the patient.⁶⁸ Dobutamine stress echocardiography is currently the most sensitive noninvasive test for cardiac disease; it measures wall motion and can detect CAV with a sensitivity and specificity of 79% and 83% respectively.⁶⁹ Possible future modalities include both pulse-wave tissue Doppler imaging and electron-beam CT. Since both modalities are noninvasive they may replace angiography as screening tools, allowing IVUS to be used in high-risk patients or those with equivocal or positive test results.

Treatment and prevention

Treatment of established CAV in humans remains limited. Encouraging research, however, has been done in small animals. For example, treatment with anti-CD154 in a rat cardiac allograft rejection model prevents acute rejection and drastically slows the development of CAV.⁷⁰ In this study, early treatment was required to inhibit CAV.⁷⁰

In clinical heart transplantation the focus remains on prevention of CAV via attenuation of adverse non-immunologic and immunologic reactions. Before transplantation, preventing endothelial injury at brain death, reducing cold ischemic time and subsequent tissue damage, and improving myocardial preservation

during storage and transportation of the graft all aid in post-transplant cardiac function and longevity. In a study on prolonged cold storage, Kevelaitis and associates⁷¹ demonstrated that longer cold ischemic times produced greater endothelial dysfunction in cardiac allografts and that the composition of the storage medium affected the extent of allograft tissue damage. Our group has shown that profound hypothermic storage results in depressed myocardial metabolic and functional recovery⁷² and that shed donor blood perfusion can permit cardiac allograft storage at tepid temperatures, resulting in improved myocardial performance.^{72,73} We have also shown that the addition of insulin to the blood perfusate during storage results in improved functional and metabolic recovery during heart transplantation.⁷⁴ Fedak and associates⁷⁵ demonstrated that bosentan, an endothelin-1 antagonist, added to shed blood perfusion improves both the functional recovery of the myocardium and endothelium. Several other groups have demonstrated that endothelin antagonism reduces CAV.^{76,77}

Immediately after transplantation, patients are placed on calcineurin immunosuppressive drugs (cyclosporine or tacrolimus), most commonly cyclosporine. Unfortunately, cyclosporine in high doses and for a long time can cause side effects such as renal failure⁴ and hypertension. Simonson and colleagues⁷⁸ demonstrated in a Lewis to Fischer rat heart transplant model that the combination of low-dose cyclosporine with an endothelin-converting enzyme inhibitor resulted in long-term survival of the graft equal to that of high-dose cyclosporine alone. As an alternative to using cyclosporine, other immunosuppressive drugs such as mycophenolate mofetil, rapamycin or leflunomide, may inhibit CAV by limiting smooth-muscle cell proliferation.⁴ The newer immunosuppressive agent everolimus has recently been demon-

strated to reduce the frequency and severity of CAV⁷⁹ in humans. Eisen and associates⁷⁹ demonstrated that, in patients on cyclosporine and corticosteroids, everolimus reduces intimal thickness and index compared with azathioprine.

Hyperlipidemia is known to be a risk factor for both CAD and CAV. Unfortunately, immunosuppressive therapy with corticosteroids, cyclosporine, rapamycin and to a lesser extent tacrolimus and everolimus results in hyperlipidemia.⁸⁰ To treat hyperlipidemia in post-transplant patients, lipid-lowering drugs are prescribed since lifestyle changes are usually not enough to lower lipid profiles to desired levels. HMG-CoA reductase inhibitors, or statins, are the most popular and are very effective at lowering total cholesterol, low-density lipoprotein (LDL) and very low-density lipoprotein, and increasing high-density lipoprotein (HDL). Recently, it has been documented that statins have pleiotropic effects in that they improve vascular function. Statins decrease endothelial dysfunction through increasing nitric oxide production, inhibiting the coagulation cascade and limiting oxidized-LDL-mediated damage to the endothelium.⁸¹⁻⁸³ Pravastatin is the most commonly used HMG-CoA reductase inhibitor after heart transplantation. In 1995, Kobashigawa and associates⁸⁴ showed that treatment with pravastatin (40 mg/d) for 1 year, lowered mean LDL and triglyceride levels, raised HDL levels and reduced intimal thickening and cardiac rejection accompanied by hemodynamic compromise ($p = 0.002$) (Fig. 2). In this trial, patients treated with pravastatin had a lower incidence of CAV and improved survival ($p = 0.025$)⁸⁴ (Fig. 3). These effects may be enhanced through immunosuppression modulation since a subgroup of patients on pravastatin had significantly reduced cytotoxicity of natural killer cells.⁸⁴ Simvastatin, likewise, has beneficial lipid-lowering effects in heart transplant recipients.⁸⁵ In addition,

Simvastatin inhibits proliferation and induces apoptosis of vascular smooth-muscle cells.⁸⁶ However, simvastatin has a low but significant rate of rhabdomyolysis and myositis. Thus, Keogh and associates⁸⁵ proposed that pravastatin be the statin of choice in heart transplantation. Atorvastatin has been shown to further reduce LDL in heart transplant recipients who are resistant to pravastatin or simvastatin. However, of the 48 patients who had received a mean (and standard deviation) atorvastatin dose of 21 (10) mg, 2 suffered from myositis, and myalgias appeared in another 2 patients. The study concluded that the drug was safe in moderate doses with careful patient monitoring.⁸⁷

HHcy in the cardiac transplant patient affects long-term outcomes by leading to the development of CAV. Several investigators have demonstrated that folic acid and vitamin B₁₂ supplementation can significantly reduce homocysteine levels in the cardiac transplant patient.⁸⁸⁻⁹⁰ Kutschka and associates⁹⁰ demonstrated that folic acid supplementation (5 mg/d) can effectively lower elevated homocysteine levels in heart transplant recipients. Unfortunately, these studies revealed only that homocysteine levels

can be lowered and did not demonstrate if reduction leads to decreased severity or prevalence of CAV.

There is general acceptance that alloimmunity plays a role in CAV. The occurrence of CAV increases as the number of HLA mismatches increases.^{91,92} Before transplantation most patients have a panel reactive antibody (PRA) test performed. A PRA result greater than 10% is considered positive and indicates that the recipient will be at higher risk for graft rejection. Kerman and associates⁹³ demonstrated that recipients with PRAs greater than 10% had a 2-fold increased risk for CAV. Immune modulation to lower PRAs has the potential to reduce acute rejection and may limit the development of CAV. Treatment strategies to lower PRAs include the intravenous use of immunoglobulin, plasmapheresis, cyclophosphamide, mycophenylate mofetil and azathioprine. The optimal strategy to prevent alloimmune injury would be to induce tolerance. Host tolerance to the allograft will abolish rejection and the immune component of CAV development. Although not achieved clinically, several investigators have demonstrated that tolerance can be induced in experimental models.⁹⁴⁻⁹⁷

Once CAV has been established,

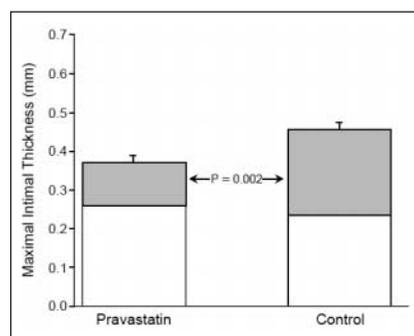


FIG. 2. Maximal intimal thickness 1 year after cardiac transplantation. Pravastatin significantly attenuated intimal proliferation during the first year after transplantation. White = baseline, grey = increase from baseline to 1 year. Reproduced with permission from Kobashigawa JA, Katznelson S, Laks H, Johnson JA, Yeatman L, Wang XM, et al. Effect of pravastatin in outcomes after cardiac transplantation. *N Engl J Med* 1995;333:621-7.

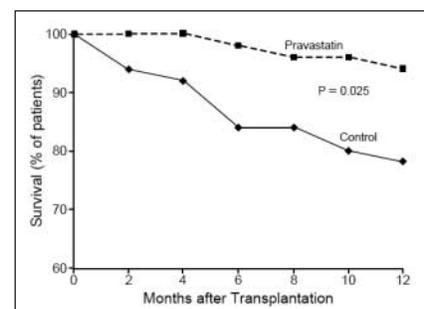


FIG. 3. Survival during the first year after cardiac transplantation. Treatment with pravastatin significantly ($p = 0.025$) improved survival compared with controls. Solid line = control, dotted line = pravastatin. Reproduced with permission from Kobashigawa JA, Katznelson S, Laks H, Johnson JA, Yeatman L, Wang XM, et al. Effect of pravastatin in outcomes after cardiac transplantation. *N Engl J Med* 1995;333:621-7.

treatments such as coronary angioplasty, coronary stenting, and coronary bypass offer only palliative solutions. The only true solution to severe CAV is repeat heart transplantation. Even so, CAV is likely to recur, and there are significant moral and ethical issues that complicate repeat transplantation.^{5,98}

Summary and conclusions

CAV is the major limiting factor for long-term survival after heart transplantation. It affects up to 75% of patients 3 years after transplantation. The risk factors for CAV can be divided in 2 categories (immunologic and nonimmunologic). Immunologic factors include the severity and frequency of acute rejection, and chronic rejection. Nonimmune factors include the classic risk factors for CAD, ischemia-reperfusion injury during organ retrieval and transplantation, CMV infection and endothelial injury from immunosuppressive drug therapy. Current areas of research focus on determining the etiology of CAV and the development of treatment strategies to prevent or limit its extent. These include endothelial protection during organ retrieval, limiting the use of calcineurin inhibitors and aggressive management of CAD risk factors. Another challenge in the management of CAV is its diagnosis. Early diagnosis of CAV will lead to earlier treatment and better outcomes. Angiography — the standard diagnostic modality for CAD — lacks sensitivity, and IVUS (the most sensitive method) lacks the ability to assess the entire coronary tree. New diagnostic tools are required for the more accurate and earlier diagnosis of CAV. The successful long-term survival of the cardiac transplant patient rests in our ability to understand, detect, treat and prevent CAV.

CAV is a multifactorial disease that remains the major limitation to long-term survival after heart transplantation. Methods of diagnosis have im-

proved significantly with the use of IVUS in addition to angiography. Since treatment of CAV is limited and usually involves repeat transplantation, prevention of immunologic and non-immunologic risk factors is of critical importance. CAV is conceptually very similar to post-transplant disorders in other organs (e.g., bronchiolitis obliterans with organizing pneumonia, biliary cirrhosis). Therefore, novel therapeutic strategies to prevent or attenuate the development of CAV may have clinical relevance to transplant recipients of other solid organs.

Competing interests: None declared.

References

- Hertz MI, Taylor DO, Trulock EP, Boucek MM, Mohacs PJ, Edwards LB, et al. The registry of the International Society for Heart and Lung Transplantation: nineteenth official report — 2002. *J Heart Lung Transplant* 2002;21:950-70.
- Labarrere CA, Nelson DR, Faulk WP. Myocardial fibrin deposits in first month after transplantation predict subsequent coronary artery disease and graft failure in cardiac allograft recipients. *Am J Med* 1998;105:207-13.
- Labarrere CA. Anticoagulation factors as predictors of transplant-associated coronary artery disease. *J Heart Lung Transplant* 2000;19:623-33.
- Gamba A, Mammana C, Fiocchi R, Iamele L, Mamprin F. Cyclosporine and graft coronary artery disease after heart transplantation. *Compr Ther* 2000;26:121-6.
- Aranda JM, Hill J. Cardiac transplant vasculopathy. *Chest* 2000;118:1792-800.
- Billingham ME. Histopathology of graft coronary disease. *J Heart Lung Transplant* 1992;11(3 Pt 2):S38-44.
- Lozano MD. Microvascular coronary arterial vasculopathy — predictive value of endomyocardial biopsy [review]. *Z Kardiol* 2000;89 Suppl 9:IX/54-7.
- Russell ME. Cardiac allograft vasculopathy — a changing perspective. *Z Kardiol* 2000;89 Suppl 9:IX/6-10.
- Johnson DE, Gao SZ, Schroeder JS, DeCampi WM, Billingham ME. The spectrum of coronary artery pathologic findings in human cardiac allografts. *J Heart Lung Transplant* 1989;8:349-59.
- Gao SZ, Alderman EL, Schroeder JS, Silverman JF, Hunt SA. Accelerated coronary vascular disease in the heart transplant patient: coronary arteriographic findings. *J Am Coll Cardiol* 1988;12:334-40.
- Hoang K, Chen YD, Reaven G, Zhang L, Ross H, Billingham M, et al. Diabetes and dyslipidemia. A new model for transplant coronary artery disease. *Circulation* 1998;97:2160-8.
- Kemna MS, Valantine HA, Hunt SA. Metabolic risk factors for atherosclerosis in heart transplant recipients. *Am Heart J* 1994;128:68-72.
- Benza RL, Tallaj J. Cardiac allograft vasculopathy (chronic rejection). In: Kirklin JK, McGiffin DC, editors. *Heart transplantation*. Philadelphia: Churchill Livingstone; 2002. pp. 615-64.
- Costanzo-Nordin MR. Cardiac allograft vasculopathy: relationship with acute cellular rejection and histocompatibility. *J Heart Lung Transplant* 1992;11(3 Pt 2):S90-103.
- Esper E, Glagov S, Karp RB, Simonsen KK, Filer SR, Scanu AM, et al. Role of hypercholesterolemia in accelerated transplant coronary vasculopathy: results of surgical therapy with partial ileal bypass in rabbits undergoing heterotopic heart transplantation. *J Heart Lung Transplant* 1997;16:420-35.
- Fodinger M, Wagner OF, Horl WH, Sunder-Plassmann G. Recent insights into the molecular genetics of homocysteine metabolism. *Kidney Int* 2001;59:S238-42.
- Fodinger M, Sunder-Plassmann G. Increased cysteine plasma levels in kidney transplants: a potential vascular disease risk factor? *Transplantation* 2001;71:713-5.
- Sunder-Plassmann G, Fodinger M. Cost-effectiveness of homocysteine-lowering therapy to prevent coronary heart disease. *JAMA* 2002;287:190.
- Stein G, Muller A, Busch M, Fleck C, Sperschneider H. Homocysteine, its metabolites, and B-group vitamins in renal transplant patients. *Kidney Int Suppl* 2001;78:S262-5.
- Ambrosi P, Garcon D, Riberi A, Habib G, Barlatier A, Kreitmann B, et al. Association of mild hyperhomocysteinemia with cardiac graft vascular disease. *Atherosclerosis* 1998;138:347-50.
- Ambrosi P, Barlatier A, Habib G, Garcon D, Kreitman B, Roland PH, et al. Hyperhomocysteinemia in heart transplant recipients. *Eur Heart J* 1994;15:1191-5.
- Langman LJ, Ray JG, Evrovski J, Yeo E, Cole DE. Hyperhomocyst(e)inemia and the increased risk of venous thromboembolism: more evidence from a case-control study. *Arch Intern Med* 2000;160:961-4.
- Cook RC, Tupper JK, Parker S, Kingsbury K, Frohlich JJ, Abel JG, et al. Effect of immunosuppressive therapy, serum creatinine, and time after transplant on plasma total homocysteine in patients following heart transplantation. *J Heart Lung Transplant* 1999;18:420-4.
- Cook RC, Parker S, Kingsbury K,

- Frohlich JJ, Abel JG, Gao M, et al. Effective treatment of hyperhomocysteinemia in heart transplant recipients with and without renal failure. *J Heart Lung Transplant* 2001;20:310-5.
25. Cole DE. Homocysteine as a risk factor in cardiovascular disease. *Adv Exp Med Biol* 2001;498:59-64.
 26. Miner SE, Cole DE, Evrovski J, Verma A, Daly PA, Ross HJ. Hyperhomocysteinemia and transplant coronary artery disease in cardiac transplant recipients. *Clin Transplant* 2001;15:258-62.
 27. Miner SE, Cole DE, Evrovski J, Forrest Q, Hutchison S, Holmes K, et al. Pyridoxine improves endothelial function in cardiac transplant recipients. *J Heart Lung Transplant* 2001;20:964-9.
 28. Cole DE, Ross HJ, Evrovski J, Langman LJ, Miner SE, Daly PA, et al. Correlation between total homocysteine and cyclosporine concentrations in cardiac transplant recipients. *Clin Chem* 1998;44:2307-12.
 29. Welch GN, Loscalzo J. Homocysteine and atherothrombosis. *N Engl J Med* 1998;338:1042-50.
 30. Doshi SN, Goodfellow J, Lewis MJ, McDowell IF. Homocysteine and endothelial function. *Cardiovasc Res* 1999;42:578-82.
 31. Chambers JC, Ueland PM, Wright M, Dore CJ, Refsum H, Kooner JS. Investigation of relationship between reduced, oxidized and protein-bound homocysteine and vascular endothelial function in healthy human subjects. *Circ Res* 2001;89:187-92.
 32. Upchurch GR Jr, Welch GN, Fabian AJ, Freedman JE, Johnson JL, Keane JF Jr, et al. Homocyst(e)ine decreases bioavailable nitric oxide by a mechanism involving glutathione peroxidase. *J Biol Chem* 1997;272:17012-7.
 33. Zhang X, Li H, Jin H, Brodsky S, Goligorsky MS. Effects of homocysteine on endothelial nitric oxide production. *Am J Physiol Renal Physiol* 2000;279:F671-8.
 34. Ambrosi P, Garcon D, Riberi A, Habib G, Barlatier A, Kreitmann B, et al. Association of mild hyperhomocysteinemia with cardiac graft vascular disease. *Atherosclerosis* 1998;138:347-50.
 35. Gupta A, Moustapha A, Jacobsen DW, Goormastic M, Tuzcu EM, Hobbs R, et al. High homocysteine, low folate, and low vitamin B6 concentrations: prevalent risk factors for vascular disease in heart transplant recipients. *Transplantation* 1998;65:544-50.
 36. Ardehali A, Fischbein MP, Yun J, Irie Y, Fischbein MC, Laks H. Indirect alloreactivity and chronic rejection. *Transplantation* 2002;73:1805-7.
 37. Game DS, Warrens AN, Lechler RL. Rejection mechanisms in transplantation. *Wien Klin Wochenschr* 2001;113:832-8.
 38. Fischbein MP, Yun J, Laks H, Irie Y, Fischbein MC, Espejo M, et al. CD8+ lymphocytes augment chronic rejection in a MHC class II mismatched model. *Transplantation* 2001;71:1146-53.
 39. Szeto WY, Krasinskas AM, Kreisel D, Krupnick AS, Popma SH, Rosengard BR. Depletion of recipient CD4+ but not CD8+ T lymphocytes prevents the development of cardiac allograft vasculopathy. *Transplantation* 2002;73:1116-22.
 40. Costanzo MR, Naftel DC, Pritzker MR, Heilman JK, Boehmer JP, Brozena SC, et al. The Cardiac Transplant Research Database: heart transplant coronary artery disease detected by coronary angiography: a multi-institutional study of preoperative donor and recipient risk factors. *J Heart Lung Transplant* 1998;17:744-53.
 41. Faulk WP, Labarrere CA, Pitts D, Halbrook H. Laboratory-clinical correlates of time-associated lesions in the vascular immunopathology of human cardiac allografts. *J Heart Lung Transplant* 1993;12:S125-34.
 42. Mehra MR, Ventura HO, Chambers R, Collins TJ, Ramee SR, Kate MA, et al. Predictive model to assess risk for cardiac allograft vasculopathy: an intravascular ultrasound study. *J Am Coll Cardiol* 1995;26:1537-44.
 43. Uretsky BF, Murali S, Reddy GS. Development of coronary artery disease in cardiac transplant patients receiving immunosuppressive therapy with cyclosporin and prednisone. *Circulation* 1987;76:244.
 44. Schutz A, Kemkes BM, Kugler C, Angermann C, Schad N, Rienmuller R, et al. The influence of rejection episodes on the development of coronary artery disease after transplantation. *Eur J Cardiothorac Surg* 1990;4:300-7.
 45. Gohra H, MacDonal TO, Verrier ED, Aziz S. Endothelial loss and regeneration in a model of transplant arteriosclerosis. *Transplantation* 1995;60:96-102.
 46. Boyle EM Jr, Lille ST, Allaire E, Clowes AW, Verrier ED. Endothelial cell injury in cardiovascular surgery: ischemia-reperfusion. *Ann Thorac Surg* 1996;62:1868-75.
 47. Byrne JG, Smith WJ, Murphy MP, Couper GS, Appleyard RF, Cohn LH. Complete prevention of myocardial stunning, contracture, low-reflow, and edema after heart transplantation by blocking neutrophil adhesion molecules during reperfusion. *J Thorac Cardiovasc Surg* 1992;104:1589-96.
 48. Tsao PS, Aoki N, Lefer DJ, Johnson G 3rd, Lefer AM. Time course of endothelial dysfunction and myocardial injury during myocardial ischemia and reperfusion in the cat. *Circulation* 1990;82:1402-12.
 49. Tsao PS, Lefer AM. Time course and mechanism of endothelial dysfunction in isolated ischemic and hypoxic perfused rat heart. *Am J Physiol* 1994;266:H128-36.
 50. Seccombe JF, Schaff HV. Coronary artery endothelial function after myocardial ischemia and reperfusion. *Ann Thorac Surg* 1995;60:778-88.
 51. Bienvenu K, Granger DN. Molecular determinants of shear rate-dependent leukocyte adhesion in postcapillary venules. *Am J Physiol* 1993;264:H1504-8.
 52. Ma XL, Weyrich AS, Lefer DJ, Buerke M, Albertine KH, Kishimoto TK, et al. Diminished basal nitric oxide release after myocardial ischemia and reperfusion promotes neutrophil adherence to coronary endothelium. *Circ Res* 1993;72:403-12.
 53. Smith CW, Entman ML, Lane CL, Beaudet AL, Ty TI, Youker K, et al. Adherence of neutrophils to canine cardiac myocytes in vitro is dependent on intercellular adhesion molecule-1. *J Clin Invest* 1991;88:1216-23.
 54. Baldwin WM 3rd, Pruitt SK, Brauer RB, Daha MR, Sanfilippo F. Complement in organ transplantation. Contributions to inflammation, injury and rejection [review]. *Transplantation* 1995;59:797-808.
 55. Day JD, Rayburn BK, Gaudin PB, Baldwin WM 3rd, Lowenstein CJ, Kasper EK, et al. Cardiac allograft vasculopathy: the central pathogenic role of ischemia-induced endothelial cell injury [review]. *J Heart Lung Transplant* 1995;14(6 pt 2):S142-9.
 56. Yamani MH, Haji SA, Starling RC, Tuzcu EM, Ratliff NB, Cook DJ, et al. Myocardial ischemic-fibrotic injury after human heart transplantation is associated with increased progression of vasculopathy, decreased cellular rejection and poor long-term outcome. *J Am Coll Cardiol* 2002;39:970-7.
 57. Subramanian AK, Quinn TC, Kickler TS, Kasper EK, Tucker PC. Correlation of *Chlamydia pneumoniae* infection and severity of accelerated graft arteriosclerosis after cardiac transplantation. *Transplantation* 2002;73:761-4.
 58. Grattan MT, Moreno-Cabral CE, Starnes VA, Oyer PE, Stinson EB, Shumway NE. Cytomegalovirus infection is associated with cardiac allograft rejection and atherosclerosis. *JAMA* 1989;261:3561-6.
 59. Weis M, Kleidal TN, Lin KY, Panchal SN, Gao SZ, Valantine HA, et al. Cytomegalovirus infection impairs the nitric oxide synthase pathway: role of asymmetric dimethylarginine in transplant arteriosclerosis. *Circulation* 2004;109:500-5.
 60. Botas J, Pinto FJ, Chenzbraun A, Liang D, Schroeder JS, Oesterle SN, et al. Influence of preexistent donor coronary artery disease on the progression of transplant vasculopathy: an intravascular ultrasound study. *Circulation* 1995;92:1126-32.
 61. Takada M, Nadeau KC, Hancock WW,

- Mackenzie HS, Shaw GD, Waaga AM, et al. Effects of explosive brain death on cytokine activation of peripheral organs in the rat. *Transplantation* 1998;65:1533-42.
62. Wilhelm MJ, Pratschke J, Beato F, Taal M, Kusaka M, Hancock WW, et al. Activation of the heart by donor brain death accelerates acute rejection after transplantation. *Circulation* 2000;102:2426-33.
63. Nissen S. Coronary angiography and intravascular ultrasound. *Am J Cardiol* 2001;87(4A):15A-20A.
64. Rickenbacher PR, Pinto FJ, Chenzbraun A, Botas J, Lewis NP, Alderman EL, et al. Incidence and severity of transplant coronary artery disease early and up to 15 years after transplantation as detected by intravascular ultrasound. *J Am Coll Cardiol* 1995;25:171-7.
65. Schoenhagen P, Nissen S. Understanding coronary artery disease: tomographic imaging with intravascular ultrasound. *Heart* 2002;88:91-6.
66. Tuzcu EM, Kapadia SR, Tutar E, Ziada KM, Hobbs RE, McCarthy PM, et al. High prevalence of coronary atherosclerosis in asymptomatic teenagers and young adults: evidence from intravascular ultrasound. *Circulation* 2001;103:2705-10.
67. Mehra MR, Ventura HO, Stapleton DD, Smart FW, Collins TC, Ramee SR. Presence of severe intimal thickening by intravascular ultrasonography predicts cardiac events in cardiac allograft vasculopathy. *J Heart Lung Transplant* 1995;14:632-9.
68. Spes CH, Angermann CE. Stress echocardiography for assessment of cardiac allograft vasculopathy. *Z Kardiol* 2000;Suppl 9:IX/50-3.
69. Spes CH, Mudra H, Schnaack SD, Klaus V, Reichle FM, Überfuhr P, et al. Dobutamine stress echocardiography for non-invasive diagnosis of cardiac allograft vasculopathy: a comparison with angiography and intravascular ultrasound. *Am J Cardiol* 1996;78:168-74.
70. Yuan X, Dong VM, Coito AJ, Waaga AM, Salama AD, Benjamin CD, et al. A novel CD154 monoclonal antibody in acute and chronic rat vascularized cardiac allograft rejection. *Transplantation* 2002;73:1736-42.
71. Kevelaitis E, Nyborg NC, Menasche P. Coronary endothelial dysfunction of isolated hearts subjected to prolonged cold storage: patterns and contributing factors. *J Heart Lung Transplant* 1999;18:239-47.
72. Rao V, Feindel CM, Cohen G, Borger MA, Boylen P, Ross HJ. Is profound hypothermia required for storage of cardiac allograft? *J Thorac Cardiovasc Surg* 2001;122:501-7.
73. Rao V, Feindel CM, Weisel RD, Boylen P, Cohen G. Donor blood perfusion improves myocardial recovery after heart transplantation. *J Heart Lung Transplant* 1997;16:667-73.
74. Rao V, Feindel CM, Cohen G, Borger MA, Boylen P, Ross HJ. Effect of metabolic stimulation on cardiac allograft recovery. *Ann Thorac Surg* 2001;7:219-25.
75. Fedak PW, Rao V, Ramzy D, Verma S, Tumiati LC, Boylen P, et al. Combined endothelial and myocardial protection by endothelin antagonism enhances transplant allograft preservation. *J Thorac Cardiovasc Surg* 2005;129:407-15.
76. Okada K, Nishida Y, Murakami H, Sugimoto I, Kosaka H, Morita H, et al. Role of endogenous endothelin in the development of graft arteriosclerosis in rat cardiac allografts. Antiproliferative effects of bosentan, a nonselective endothelin receptor antagonist. *Circulation* 1998;97:2346-51.
77. Yamaguchi A, Miniati DN, Hirata K-I, Hoyt G, Robbins RC. Ex vivo blockade of endothelin-1 inhibits graft coronary artery disease in a rodent cardiac allograft model. *J Heart Lung Transplant* 2002;21:417-24.
78. Simonson MS, Robinson AV, Schulak JA, Hricik DE. Inhibition of endothelin-1 improves survival and vasculopathy in rat cardiac transplants treated with cyclosporine. *Transplantation* 2002;73:1054-9.
79. Eisen HJ, Tuzcu EM, Dorent R, Kobashigawa J, Mancini D, Kaepler HA, et al. Everolimus for the prevention of allograft rejection and vasculopathy in cardiac-transplant recipients. *N Engl J Med* 2003;349:847-58.
80. Fellström B. Impact and management of hyperlipidemia posttransplantation. *Transplantation* 2000;70:S51-7.
81. Farmer JA. Pleiotropic effects of statins [review]. *Curr Atheroscler Rep* 2000;2:208-17.
82. Koh KK. Effects of statins on vascular wall: vasomotor function, inflammation, and plaque stability. *Cardiovasc Res* 2000;47:648-57.
83. Li D, Chen HJ, Metha JL. Statins inhibit oxidized-LDL-mediated LOX-1 expression, uptake of oxidized-LDL and reduction in PKB phosphorylation. *Cardiovasc Res* 2001;52:130-5.
84. Kobashigawa JA, Katznelson S, Laks H, Johnson JA, Yeatman L, Wang XM, et al. Effect of pravastatin in outcomes after cardiac transplantation. *N Engl J Med* 1995;333:621-7.
85. Keogh A, Macdonald P, Kaan A, Abooun C, Spratt P, Mundy J. Efficacy and safety of pravastatin vs simvastatin after cardiac transplantation. *J Heart Lung Transplant* 2000;19:529-37.
86. Sindermann JR, Fan L, Weigel KA, Troyer D, Muller JG, Schmidt A, et al. Differences in the effects of HMG-CoA reductase inhibitors on proliferation and viability of smooth muscle cells in culture. *Atherosclerosis* 2000;150:331-41.
87. Patel DN, Pagani FD, Koelling TM, Dyke DB, Baliga RR, Cody RJ, et al. Safety and efficacy of atorvastatin in heart transplant recipients. *J Heart Lung Transplant* 2002;21:204-10.
88. Parisi F, Danesi H, DiCiommo V, Fina F, Giannone G, Colistro F, et al. Treatment of hyperhomocysteinemia in pediatric heart transplant recipients. *J Heart Lung Transplant* 2003;22:778-83.
89. Miriuka S, Langman LJ, Kereen ES, Miner SE, Mamer OA, Delgado DH, et al. Effects of folic acid fortification and multivitamin therapy on homocysteine and vitamin B₁₂ status in cardiac transplant recipients. *J Heart Lung Transplant* 2004;23:405-12.
90. Kutschka I, Pething K, Strüber M, Dieterich C, Harringer W, Haverich A. Homocysteine — a treatable risk factor for allograft vascular disease after heart transplantation? *J Heart Lung Transplant* 2001;20:743-6.
91. Valentine H. Cardiac allograft vasculopathy after heart transplantation: risk factors and management. *J Heart Lung Transplant* 2004;23:S187-93.
92. Vassalli G, Gallino A, Weis M, von Scheidt W, Kappenberger L, von Segesser LK, et al. Alloimmunity and nonimmunologic risk factors in cardiac allograft vasculopathy. *Eur Heart J* 2003;24:1180-8.
93. Kerman RH, Susskind B, Kerman D, Lam M, Gerolami K, William J, et al. Comparison of PRA-STAT, sHLA-EIA, and anti-human globulin-panel reactive antibody to identify alloreactivity in pretransplantation sera of heart transplant recipients: correlation to rejection and posttransplantation coronary artery disease. *J Heart Lung Transplant* 1998;17:789-94.
94. Orloff MS, DeMara EM, Coppage ML, Leong N, Fallon MA, Sickle J, et al. Prevention of chronic rejection and graft arteriosclerosis by tolerance induction. *Transplantation* 1995;59:282-8.
95. Madsen JC, Yamada K, Allan JS, Choo JK, Erhorn AE, Pins MR, et al. Transplantation tolerance prevents cardiac allograft vasculopathy in major histocompatibility complex class I-disparate miniature swine. *Transplantation* 1998;65:304-13.
96. Yamada K, Choo JK, Allan JS, Erhorn AE, Menard MT, Mawulawde K, et al. The effect of thymectomy on tolerance induction and cardiac allograft vasculopathy in a miniature swine heart/kidney transplantation model. *Transplantation* 1999;68:485-91.
97. Zhang QW, Tomita Y, Matsuzaki G, Shimizu I, Iwai T, Okano S, et al. Heart allograft tolerance without development of posttransplant cardiac allograft vasculopathy in chimerism-based, drug-induced tolerance. *Transplantation* 2002;73:652-6.
98. Miniati DN, Robbins RC. Heart transplantation: a thirty year perspective. *Annu Rev Med* 2002;53:189-205.